NEW STANDARDIZED CHROMOGENIC ASSAYS FOR AUTOMATED MEASUREMENTS OF FIX OR FIXa IN PLASMA AND THERAPEUTIC CONCENTRATES

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INTRODUCTION

- Automated measurements of Factor IX (FIX) and activated Factor IX (FIXa) are required for testing Prothrombin Complex Concentrates (PCC), or therapeutic recombinant Factor IX (eg. BeneFIX®), but also for quantitating plasma Factor IX (Haemophilia B), or for performing recovery studies in treated patients.
- Two fully homogeneous chromogenic assays, highly sensitive, and offering an extended working range, were developed for these applications.

TESTED SAMPLES

- Normal citrated plasma (NI), and plasma from dicoumarol treated patients (VKA), and Haemophilia B patients (Haem. B).
- FIX deficient plasma (FIX DP), immunodepleted (FIX <0.1%).
- FIX pharmaceutical concentrates (BeneFIX®; recombinant FIX), are pre-diluted in FIX DP.
- FIX concentrations (BenefIX®, recombinant FIX), are pre-diluted in FIX DP or in the assay diluent (this predilution is made at 1 IU/ml for the high range, or 0.2 IU/ml for the low range), then assayed at the standard assay dilution for the high (1:80 and further) or low (1:15) ranges, in the assay diluent and tested with the automated instrument STA-R (Diagnostica Stago).

RESULTS

- FIX chromogenic assay dose response curves for the High and the low range (STA-R).
- FIX chromogenic assay dose response curve (STA-R).
- Compared reactivity and recovery of purified (h)FIX or two therapeutic preparations.

DISCUSSIONS

- FIX chromogenic assay (STA-R):
  - High range: 0.025 IU/ml (125 ng/ml) to 0.0006 IU/ml (3 ng/ml) in diluted sample (1:80).
  - Low range: detection threshold of 0.005 IU/ml in plasma.
  - Excellent correlation with conventional FIX clotting method (r²=0.93).
  - No interference of other factors or heparin.
  - Using the specific assay diluent, purified human FIX or two FIX pharmaceutical concentrates yield a similar reactivity when than when diluted into FIX DP (80-105% recovery).

Factor IXa chromogenic assay:
- Dynamic range: 0.025 to 0.0005 IU/ml FIXa (0.001 IU ~1 ng) (tested at 1:2 dilution).
- Sensitive and efficient for quantitating trace amounts of FIXa in FIX concentrates preparations (data not shown).

Further studies are in progress on Haemophilia B patients and various pharmaceutical preparations.

MATERIAL AND METHODS

- FIX and FIXa (Act) chromogenic assays (based on FIXa generation), designed with the use of highly purified human proteins, and well characterized synthetic phospholipids, and including an optimized diluent for full expression of FIX or FIXa activity:
  - Tested FIX is incubated with FXla, FX, FVIII:C, Fila, phospholipids and calcium.
  - FIX is then activated to FIXa, which activates FX to FIXa in a dose-dependent manner.
  - FXa activity is measured with a chromogenic substrate (405 nm).

- FIXa (Activated FIX) is measured using the same principle, but without the FIX activation step by FXla.
- Conventional aPTT based FIX clotting assay (FIX DP and Cephen aPTT reagent).
- Assay calibrations are standardized with the NIBSC standards for plasma FIX (SSC/STH secondary plasma standard lot 3) or activated FIX (97/562).

CONCLUSIONS

- Standardized assays (NIBSC), designed with optimized and secured raw materials, highly stable (72h at 2-8°C, or frozen) and fully automatable.
- These two chromogenic assays are easily performed on automated coagulation instruments and allow measuring FIX or FIXa activity with high sensitivity.
- No FIX deficient plasma is required for testing FIX or FIXa, and concentrates can be assayed directly diluted in the assay diluent.
- These assays represent a very helpful alternative to clotting methods for testing FIX in plasma, or the activation grade of PCC and other FIX therapeutic concentrates. These methods improve the laboratory practice for monitoring FIX preparations, but also for testing FIX in haemophilias or monitoring recovery of therapeutic concentrates in patients receiving substitutive therapy.
New standardized chromogenic assays for automated measurements of Factor IX or Factor IXa in plasma and therapeutic concentrates.

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Automated measurements of Factor IX and Factor IXa are required for testing Prothrombin Complex Concentrates (PCC), plasma Factor IX (Haemophilia B) or recombinant Factor IX (BenefIX), or for performing recovery studies in treated patients. Two chromogenic assays were developed for measuring Factors IX or IXa in plasma or in concentrates. For quantitating Factor IX, diluted specimen is incubated with a constant and in excess amount of Factor X, phospholipids, calcium, and Factor VIII:C. Factor IX activation is initiated by Factor Xla containing human thrombin (necessary for Factor VIII:C activation). There is a direct relationship between Factor IX in the tested sample and Factor Xa generation. Factor Xa is then measured with a specific chromogenic substrate. Finally, the colour development, measured at 405 nm, is a direct relationship of Factor IX concentration. The assay is standardized with the International Standards for plasma Factor IX or Factor IX concentrates (NIBSC). The dynamic range in the assayed dilution is from 0.02 IU/ml (100 ng/ml) to 0.0005 IU/ml (2.5 ng/ml). Plasmas are assayed diluted 1:100. A low range is available for Factor IX in Haemophiliacs, with a detection threshold of 0.005 IU/ml in plasma. The assay presents an excellent correlation with conventional clotting methods in normals, dicoumarol treated patients or B Haemophiliacs ($r^2=0.93$). There is no interference of other factors in the tested specimen. No Factor IX deficient plasma is required for testing Factor IX or Factor IXa, and concentrates can be assayed directly diluted in the assay diluent. Factor IXa is measured with a similar method, omitting the activation by Factor Xla. The dynamic range is from 0.025 IU/ml to 0.0005 IU/ml of Factor IXa (0.001 IU is about 1 ng). This assay is standardized with the International Standard for Factor IXa (NIBSC). It is very helpful for testing the activation grade of PCC and other Factor IX therapeutic concentrates. Automation can be easily performed for both assays on all the automated coagulation instruments. These methods improve the laboratory practice for monitoring Factor IX preparations.