Matched-Pair Antibody Set
for ELISA of human
von Willebrand Factor antigen (vWF)

Sufficient reagent for **5 x 96 well** plates

Product #:	VWF-EIA
Lot #:	SAMPLE
Expiry Date:	SAMPLE

Store at –10 to -20°C

For Research Use Only
Not for use in diagnostic procedures.

Description of von Willebrand Factor (vWF)

von Willebrand Factor (vWF, also previously referred to as Factor VIII related antigen) is a large adhesive protein produced in endothelial cells and megakaryocytes. There are two critical functions of vWF, the first being its involvement in the process of platelet adhesion and aggregation through interaction with platelet receptor glycoprotein Ib, the second being the binding and stabilization of Factor VIII (antihaemophilic factor) for secretion and transport in plasma. The vWF precursor protein is synthesized with a 95,000 dalton propeptide (also known as vWF antigen-II), believed to be involved in the intracellular multimerization of the vWF subunits. The mature vWF multimers are then packed into storage organelles within the cell (Weibel-Palade bodies) after which the propeptide is cleaved and released. vWF circulates as multimers of disulphide linked 220,000 Dalton subunits and the molecular weight of these multimers ranges from 0.5-20 million daltons.
The plasma concentration of vWF is typically 10 µg/ml, but increased levels are often observed in pregnancy and other conditions of physiological stress. von Willebrand's disease (vWD) is perhaps the most common inherited bleeding disorder in humans and is the result of either quantitative deficiencies of vWF (vWD Types I & III), or one of a number of qualitative disorders of vWF structure and function (vWD Type II) [1-3].

Principle of Sandwich-style ELISA

Affinity-purified antibody to vWF is coated onto the wells of a microtitre plate. Any remaining binding sites on the plastic wells are blocked with an excess of bovine serum albumin. The plates are washed and plasma or other fluids containing vWF are applied. The coated antibody will capture the vWF in the sample. After washing the plate to remove unbound material, a peroxidase conjugated second antibody to vWF is added to the plate to bind to the captured vWF. After washing the plate to remove unbound conjugated antibody, the peroxidase activity is expressed by incubation with o-phenylenediamine (OPD). After a fixed development time the reaction is quenched with the addition of H_2SO_4 and the colour produced is quantified using a microplate reader. The colour generated is proportional to the concentration of vWF present in the sample.

Supplied Materials:

1. Capture Antibody (VWF-EIA-C): One yellow low-capped vial containing 0.5 ml of polyclonal purified anti-vWF antibody for coating plates.

2. Detecting Antibody (VWF-EIA-D): One red-capped vial containing 0.5 ml of peroxidase conjugated polyclonal anti-vWF antibody for detection of captured vWF.

Note: Antibodies are supplied in a 50% (v/v) glycerol solution for storage at –10 to -20°C. Keep vials tightly capped. Do not store in frost-free freezers.

Materials Required but not Provided:

1. Coating Buffer: 50 mM Carbonate
1.59g of Na_2CO_3 and 2.93g of $NaHCO_3$ up to 1 litre. Adjust pH to 9.6. Store at 2-8°C up to 1 month.

2. PBS: (base for wash buffer and blocking buffer)
8.0g NaCl, 1.15g Na_2HPO_4, 0.2g KH_2PO_4 and 0.2g KCl, up to 1 litre. Adjust pH to 7.4, if necessary. Store up to 1 month at 2-8°C, discard if there is evidence of microbial growth.

3. Wash Buffer: PBS-Tween (0.1%,v/v)
To 1 litre of PBS add 1.0 ml of Tween-20.
Check that the pH is 7.4. Store at 2-8°C up to 1 week.

4. Blocking Buffer: PBS-BSA (1%, w/v)
Dissolve 2.5 g of Bovine Serum Albumin (Sigma-RIA grade) in 200 ml of PBS. Adjust pH to 7.4, if required, then make up to 250 ml with PBS. Aliquot and store frozen at -20°C.

5. Sample Diluent: HBS-BSA-T20
5.95g HEPES (free acid), 1.46 g NaCl, 2.5 g Bovine Serum Albumin (Sigma, RIA grade) dissolved in 200 ml H_2O. Add 0.25 ml of Tween-20, check and adjust pH to 7.2 with NaOH, then make up to a final volume of 250 ml with H_2O. Aliquot and store frozen at -20°C.

6. Substrate Buffer: Citrate-Phosphate buffer pH 5.0
2.6g Citric acid and 6.9g Na_2HPO_4 up to a final volume of 500 ml with purified H_2O. Store at 2-8°C up to 1 month.

7. OPD Substrate: (o-Phenylenediamine.2HCl) TOXIC!
(5mg tablets: Sigma # P-6912). Make up immediately before use. Dissolve 5mg OPD in 12 ml substrate buffer then add 12 µl 30% H_2O_2. Do not store.

8. Stopping Solution: 2.5 M H_2SO_4
Caution: VERY CORROSIVE! GENERATES HEAT ON DILUTION! Where stock sulphuric acid is 18 Molar, add 13.9 ml to 86 ml H_2O. Store at room temperature.

9. Other:
Microplates, 96-well Immulon 4-HBX (http://www.labsystems.fi)
Microplate washer (optional)
Microplate reader.

Assay Procedure:

1. Coating of plates:
Dilute the capture antibody 1/100 in co ating buffer (preferably in a p olypropylene tube) and immediately add 100 µl to every well in the plate.
Incubate 2 hrs @ 22°C or overnight at 2-8°C.

2. Blocking:
Empty contents of plate and add 150 µl of blocking buffer to every well and incubate for 60 minutes @ 22°C.
Wash plate X 3 with wash buffer.

3. Samples:
Reference plasma is diluted 1/100 (100%) then serial 1/2's down to 1/3200 (3.13%). S ample plasmas are diluted 1/200, 1/400 & 1/800. All dilutions are made in HBS-BSA-T20 sample diluent. Apply 100 µl/well and incubate plate @ 22°C for 90 minutes. Wash plate X 3 with wash buffer.

4. Detecting Antibody:
Dilute the detecting antibody 1/100 in H BS-BSA-T20 sample diluent and apply 100 µl to each well. Incubate plate @ 22°C for 90 minutes. Wash plate X 3 with wash buffer.

5. OPD Substrate:
Apply 100 µl of freshly prepared OPD substrate to every well. Allow colour to develop for 10-15 minutes then stop colour reaction with the addition of 50 µl/well of 2.5 M H_2SO_4. The plate can be read at a wavelength of 490 nm.

Calculation of Results:

The construction of a p roper reference curve is of no less importance than any other aspect of the assay. A reference curve should be constructed by plotting the known concentration of standards versus absorbance. This can be done manually using graph paper, or by using curve-fitting computer software. In our experience, the dose response curves of most immunoassays tend to be sigmoid in shape. Although linear regions can be identified within the curve, the best o verall fit is of ten obtained using an algorithm that provides a weighted theoretical model of fit throughout the entire curve, such as a 4-pa rameter or 5-parameter logistic curve fit [4,5]. In general, the simplest model that defines the concentration-response relationship should be used [6].
The "back-fit" test is a simple and reliable method to determine if a curve-fitting method is a ppropriate. In this test, the apparent concentrations for the absorbance values of each standard point are read from the reference curve. The derived values are compared to th e assigned values. An appropriate curve fitting method will produ ce derived valu es that closely match assigned values throughout the range of the curve, within user-defined limits[6]. The coefficient of determination (R^2) is a valuable indicator of the overall fit, but should not be used by itself in the selection of a curve fitting method, as a poor fit in a p articular region of the curve may not be evident from this value alone [5,6].
In the quality control of this product we have determined that under the conditions described above, a reference curve that is constructed using serial dilutions of normal pooled plasma, will produce a correlation coefficient (R^2) of at least 0.980 using a log-log fit, and an R^2 of at least 0.990 using a 4-parameter logistic curve fit algorithm. However, the performance characteristics of in-house assays developed using this product in other laboratories may vary slightly from ours. Different curve fitting methods may be employed but we recommen d that the back-fit test be applie d as evidence that the fitting method is appropriate.

Technical Notes:

- This paired antibody product is intended to facilitate the end user in establishing an in-house immunoassay for research purposes only. It must not be used for diagnostic applications. Assay validation is the responsibility of the end user and should be done according to user-defined protocols[6].
- Reference calibrators should be of the same matrix and anticoagulant as the samples to be tested (example serum or plasma, citrate or EDTA)
- Do not use samples diluted less than 1/10, as falsely high readings may result.
- The optimal colour development time should be determined empirically as the time required to obtain an absorbance of at le ast 1.000 at 490 nm fo r the 100% reference point, not to exceed 20 minutes.
- Rheumatoid factor in samples may interfere in ELISA by binding to the capture and/or detecting antibodies.
- The wells should not be allowed to become dry. Keep plate covered or in a humid chamber during incubations.
- Antibodies are supplied in a 50% glycerol solution and can be centrifuged briefly in a micro-centrifuge to gather residual reagent from the cap and walls of the tube.

References:

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3. Bartlett A, Dormandy KM, Hawkey CM, Stableforth P, Voller A; Factor VIII-related antigen: Measurement by Enzyme Immunoassay. British Medical Journal, 1:994-996, 1976.
4. Nix,B, Wild D, in Immunoassays, A Practical Approach, editor J.P. Gosling, pp. 239-261, Oxford University Press, 2000.
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6. FDA Guidance for Industry. Bioanalytical Method Validation; May 2001, available on the internet: www.fda.gov/cder/guidance/index.htm